

Visualization ata

Data Visualization: The Why and How



Soud Al Kharusi & Ahmad Mahmood & YOU



Blinking of Fluorescence Dyes as an Intracellular Biosensor: A k-Space Image Correlation Spectroscopy Approach

Off

Ø

In ()

out's

Oil Immersion

Oberchise NA - 1.49

100x Mag



Motivation

Many intracellular processes involve the transport of biomolecules across organelles. Understanding complex biomolecular pathways requires techniques that probe transport dynamics and local intracellular conditions. The environment-dependent photophysics of fluorescent dves offers an avenue for in vivo biosensing.5 k-space image correlation spectroscopy is an ensemble measurement technique for photophysics and transport dynamics of subdiffractive molecules. This project aims to study the On $\overline{+}$ dependence of the blinking rates of fluorescently labelled biomolecules on their environment. This is investigated via simulation and Cy5 dye in the presence of photostabilizing agents. Furthermore, a microfluidic device was designed to mimic cellular compartmentalization for future works.

k-Space Image Correlation Spectroscopy

k-space image correlation spectroscopy temporally correlates spatially Fouriertransformed image ROIs from an image series, yielding the reciprocal space correlation function, r(k, r). Dynamic quantities, including blinking and transport, of the fluorescently labelled species can be extracted from $r(k, \tau)$. Ensemble measurement allows for rapid convergence of the correlation function, minimizing the requirement for long acquisition times and temporal sampling.

Real Space vs k-Space Visualization



Temporal Autocorrelation of k-Space Image Series 1) Acquire image series 2) Perform 2D spatial 3) Perform autocorrelation and





Blinking Simulation via Gillespie Algorithm

A simulation of blinking dyes undergoing diffusion was developed to validate the kICS image analysis. The Gillespie algorithm was used to model stochastic fluorophore blinking. A two compartment simulation model was analyzed with kICS.



Instrumentation: TIRFM



just past the critical angle. An exponentially decaying evanescent wave is produced at the point of internal reflection. The fluorescence is collected by an sCMOS camera. The camera frame rate limits the temporal resolution of the image series.

Experimental Method: Microfluidics

A microfluidic device, compatible with total internal reflection fluorescence microscopy, was designed to replicate transport between intracellular compartments. The laminar flow present in microfluidics was utilized to generate two compartments that the fluorophore dye can cross via diffusion.

Design Validation: Simulations

designed in Eusion360.

Simulations were used to analyze flow within device designs. The following pipeline was developed and implemented:





3) Python simulation of

particles flowing in device with imaging parameters.

Device Fabrication: Stereolithography A rapid prototyping cost-effective microfluidic using 3D resin printing was developed.



Results Blinking Simulation: Biological Compartmentalization

Simulated diffusing fluorophores through compartments with different blinking. The blinking rates returned from shifting the kICS ROI (in red) across the image had an increased precision of boundary localization compared to an intensity profile (in blue) The diffusion coefficient recovered was 0.03 + 0.01 μ m²/s (input was 0.01 μ m²/s).



Microfluidic Device: Simulated Compartmentalization The microfluidic device made was compatible with TIRFM and generated a clear boundary between two compartments



Microfluidic device with green food dye and water flowing, illustrating the sharn boundary.

Flow within the microfluidic device Boundary generated in device with being imaged with 488 nm total water and 200 nm fluorospheres internal reflection microscony solution. Ex/Em 505/515 nm at 10 mW 33 ms exposure

Recovering Blinking Rates of Cy5 in Different Conditions

The blinking rates of a Cv5 dve attached to DNA nanotubes, in the presence of BME (Bmercaptoethanol), with and without a triplet state quencher (ergothioneine) were recovered. The off rate significantly decreased in the presence of ergothioneine, supporting that it increases the photostability of Cy5.



Future Directions



Acknowledgements

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References

1) Kein (1) L.: Kens (1): Wiseman (2) W. K.Space Image Correlation Spectroscopy: A method for accurate transport Dom, D. C., Polin, D., Freeman, P. V. K. Sakoti mage Correlation Specificscopy. In Include architect of approximation of flucrophore Photophysics. *Biophysical Journal* 2005, 97, 3061–3075
Sahayek, S., Gill, Y., Glembockete, V., Brandao, H. B., Francois, P., Cosa, G., Wiseman, P. W. A high throughput image correlation method for rapid analysis of Biorophore photoblinking and photobleaching rates. ACS Nano 2010, 13, 11955-11966



1) CAD design 3D printed. 2) Resin print post-processing to ensure PDMS will cure.

3) PDMS molding and attachment to microscope coverslip.

enetration

Buffer 2 + Dw

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Buffer 2 + Dw

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to ensure PDMS will cure.



















From: Phys. Rev. C 97.065503

Graph 1.1 - Insets







Spectre d'extinction optique (disque de 130 nm)





Spectre d'extinction optique (disque de 130 nm)

















STEADY







STEADY

From: https://clauswilke.com/dataviz/aesthetic-mapping.html

Data Viz 101





Overview



- Common things to think about
- Types of plots, and when to use them
- Trendlines vs fits
- Visualizing Uncertainties
- Schematics
- Plots that are meant for: papers, posters, talks...
- Exercises

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• Common colour blindness

red green, blue/green, yellow/red

• Colour palettes: <u>https://colorhunt.co/</u>



STEADY

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2 colours, relation?



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3 colours, relation?



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STEADY

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4 colours, relation?



STEADY

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monochromatic, relation?





- Heatmap colour palettes
 - single **pole** vs linear & monochromatic







MyMap = [zeros(256,1),linspace(0,1,256)', zeros(256,1)]



Memorable Colour Scales





Photon hit patterns of Cherenkov light from relativistic muons on cylinder using the viridis colour scale

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Overview



- Common things to think about
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To Bar or Not to Bar





Distributions!

Histograms, Kernel Densities, Boxes, Violins et al.





Histograms: Binning






Histograms vs PDFs/Kernel Density





From: https://clauswilke.com/dataviz/histograms-density-plots.html

Stacked Histograms





Overlaying Histograms/PDFs







Box Plots



Box vs Violin Plot





Violins with Strip Chart





Smoothed Violin Plot





Smoothed Violin Plot





Changing "bandwidth" of kernel. I.e. number of σ to "smear by" can make things unphysical

Surface vs Contour





Surface vs Contour





Isosurface vs Maximum Intensity





Isosurface vs Maximum Intensity





Contours

Contours can show

probability density contained within a region when used on scatter plots

or

isosurfaces (levels) of 3D surfaces





Multidimensional Scatter Plots





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Trendlines vs Fits





Trendlines vs Fits





Trendlines vs Fits: Summary



- When doing a fit, make sure to **show equations and fit results** somewhere!
- When needing a trendline to "guide the eye" either
 - Connect the dots by straight lines
 - Do some interpolation, e.g. cubic spline fit, but **state this!**
 - Consider showing the "confidence band" as a shaded region

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https://clauswilke.com/dataviz/visualizi ng-uncertainty.html#frequency-framing



• When using error bars, you need to state what they are!

















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Schematics: how to make them









Schematics: AutoCAD







Schematics: CAD





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File Formats

Different formats are meant for different media





From: https://clauswilke.com/dataviz/image-file-formats.html

File Formats

Different formats are meant for different media



Examples:

- .png are lightweight, good for online plots (load time)
- .pdf/.svg/.eps are vectorized ("infinite" zoom! (*)
- .jpg are generally not great, avoid except for compressed photographs
 - But, when *taking* photos (camera data) use .raw or .tif/.tiff file formats!

File Formats

General rule when making plots and saving them



save high-dpi (600+) .png files, or a pdf

convert and lower resolutions as needed and when necessary

Examples:

- .pdf/.eps for journals
- .svg for your website
- .png for slide presentations

Common Tripping Hazards



- Choose the **right type of plot**
 - What is it about your data that you are trying to get across?
- Choose your colours wisely
 - What is the **relationship** between the data products you are showing?
- Tick marks & axes labels
 - always go bigger than the default size
- Pick a plotting package you like, and get good at it \rightarrow set custom defaults
 - e.g. matplotlib style files (.mplstyle)

Common Tripping Hazards 2

- Will the plot be printed out?
 - Are your lines/points distinguishable in grayscale?
- Choose the right file type for exporting
 - .pdf/.eps for papers, .png/.svg for web
- Always specify what your "error bars" are
 - \circ Are they standard errors on mean? std, σ ? Or some confidence interval?
- Don't overuse Jupyter Notebooks
 - Good for plotting, not good for coding...



Acknowledgements



Thank you to everyone who contributed data and/or plots & posters to make this workshop happen!

Catherine B., Rodrigo P., Adam C., Lauren P., Gaspard B.





- 1. Open the onedrive link on Slack **#data-visualization** or the indico page
- 2. Download the **.csv** file
- 3. Try plotting the data
- 4. Upload your plots back to the onedrive!

Exercise





Exercise





Exercise





Some Resources



Data Visualization:

• Free crash-course book on data viz: <u>https://clauswilke.com/dataviz/index.html</u>

Posters:

• Audience should be able to get these items *without* you being present

• Topic

- Main method / workflow
- Conclusion
- Check out <u>#betterposter on YouTube</u>

Fantastic Software Packages & Where to Find Them

- Python: Matplotlib, seaborn
 - Consider editing your default <u>.mplstyle file</u>
- Matlab: (basically matplotlib)
- Web pages: plotly (javascript)



